

Carbon Nanomaterials and Applications in Sensors-A Review

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ABSTRACT: This is a brief review covering the recent developments on the carbon nanostructured materials-based detectors covering recently revealed works is presented. Number of works addressing experimental and theoretical knowledge are reviewed and discussed. The results for carbon nanotubes and hybrid carbon-nanostructured devices that show sensing properties in several fields were thought of for the discussions. The goal of this paper was to focus on sensor mechanisms, and the best results reached up to the current situation are making bases for more applications. Keywords: Nanomaterials, Carbon Nanotubes,

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I. INTRODUCTION

Recent advancements within the field of applied science have created opportunities for the great achievements in biosensing. Emergence of recent materials and their hybrids have raised hopes for development of new biosensors capable of delivering on site results of variety of analytes (at smaller level) at the same time without requiring skillful personnel or refined machinery. A biosensor could be a device consisting of a biological sensing element i.e the bioreceptor which detects the analyte, is connected to a detector or transducer, which converts the response into a measurable signal. On the basis of the styles of bioreceptors, biosensors will be classified as immunobiosensors, enzymatic biosensors, genobiosensors and based mostly upon the transduction process, biosensors can additionally classified he as optical, piezoelectric, electrochemical, and thermometric biosensors.

Nowadays, the need to develop new sensors with more specific properties is increasing. There are avariety of sensors for monitoring gases, heavy metals, moisture, biomolecules, pressure, etc. However, most of them are expensive, require pre-treatment, are difficult to use, slow to respond, and do not have the ideal limit of detection, Sensitivity or Selectivity. With a view to improve the above parameters, nanotechnology has provided the most promising update in material properties and has given significant advances in overcoming the limitations conventional materials once faced.

Carbon nanotubes (CNTs), as one of the extensively used 1D nanomaterials, have been applied for the fabrication of numerous highoverall performance sensors and biosensors because of the unique mechanical, electrical, and magnetic properties of CNTs. In addition, the high surface area and high adsorption capacity toward numerous molecules/biomolecule of CNTs make CNTs very good option to manufacture chemical and bio sensors with high sensitivity and selectivity. Besides CNTs, carbon nanofibers (CNFs) have additionally been broadly studied because of their unique chemical and physical properties and comparable structure to fullerenes and CNTs.

This report presents the latest advancements in the field of nanotechnology and it's application with the scope of sensors. This report also deals with the different types of the sensors that can be fabricated as an option. Recent studies will be cited from here.

II. NANOMATERIALS SUITABLE FOR USE

2.1 Carbon Nanotubes (CNTs)

CNTs are sp2 hybridized carbon atom rolled graphene sheets having hollow interior with diameter in nanosize and the length varies from nanometers to microns. Due to their excellent structural, electronic and mechanical properties, CNTs have gained huge interest from the time of their discovery. These showcase variable conductivity and feature small size which lead them to an excellent choice for use within the scope of molecular electronics as molecular wires. Functionalization of CNT may be performed with the aid of using both covalent and non-covalent



binding with different chemical groups which makes CNTs biocompatible for conjugation with biomolecules and hence making them a appropriate candidate for biosensing. The functional groups which include amine and carboxyl, increase the rate of electron transfer. In practical, the watersoluble polymers based functionalization of CNTs or surface functionalization with ionic or hydrophilic groups of CNTs allows to attain CNT solubilization in aqueous media and is a giant parameter for CNTs to work as a assisting matrix or scaffold for the entrapment of proteins/DNA/ Antibody/enzymes. Functionalization of CNTs additionally improves the direct electron transfer among the biological element active sites and the electrode. Because of the above reason, conductivity of CNTs (especially MWCNTs) is normally stepped forward with the aid of using functionalizing them with redox polymers, hapten molecules, thiol derivatives and N-ethyl-N-(3carbodimide-N-hydroxy dimethylaminopropyl succinimide (EDS-NHS). An amperometric acrylamide biosensor was fabricated with the aid of using immobilization of Hb onto nanocomposite of cMWCNT and Fe2O4 nanoparticles (NPs) electrodeposited onto gold electrode via a polymer chitosan film (Batra et al., 2013). A great improved sensitivity and selectivity was acquired for acrylamide. The sensor offered a short response, wider linear range, decreased limit of detection (0.02 nM), excellent reproducibility, and long stability. Recently, an immunosensor was mentioned for cancer detection making use of poly (diallyldimethylammonium chloride) (PDDA)functionalized CNTs for assembly of HRP and concanavalin A (ConA) at the gold electrode. Utilizing the biospecific interaction among HRP and ConA, PDDA changed CNTs form a complex of CNTs/PDDA/HRP/ConA which afterward mixed with Ab labeled HRP and form a sandwich. The immunosensor shows a good linear range from 0.05-5 ng/mL to 5-200 ng/mL and a detection restriction of 0.018 ng/mL (Yang et al., 2014). A fresh glucose biosensor was fabricated by the use of free CNTs and enzymatic electrode immobilizing glucose oxidase (GOx), GOx coating and GOx precipitate coating. This sensor contemplated excellent development in sensing, stability and electron transfer (Kim et al. 2015).

2.2 Pure Carbon NanoFibers(CNFs)

Due to their high specific surface area and good electrocatalytic ability towards the oxidation of specific organic matter, pure CNFs are normally preferred to observe small molecules, viruses, proteins, and nucleic acids in food quality check and clinical analysis. For example, yue et al. said mesoporous CNF-modified pyrolytic graphite electrode for the cooccurring determination of uric acid, ascorbic acid, and dopamine. Koehn et al. made a vertically aligned CNF electrode array by the PECVD method, then integrated the CNF array with the wireless instantaneous neurotransmitter detector system to detect dopamine by quick scan cyclic voltammetry. Rand and coworkers developed a biosensor supported vertically aligned CNFs for the cooccurring detection of serotonin and dopamine within the presence of excess ascorbic acid. Periyakaruppan et al. says similar CNFs primarily based nanoelectrode arrays for label-free detection of cardiac troponin-I in the early diagnosing of heart muscle infarction.

2.3 CNFs modified with metal oxides

Since some acid gases and organic gases will cause changes within the impedance of metal oxide-decorated CNFs, metal oxide-decorated CNFs will be used for the detection of specific acid gas and organic gas. Lee and coworkers made ZnO/SnO2 nanonodules-decorated CNFs for dimethylphosphonate gas detection by single nozzle co-electrospinning by 2 phase-separated polymer solutions. Later, this group changed WO3 nanonodule to the surface of CNFs for the detection of NO2 gas using an equivalent method, and observed that the sensitivity of the WO3 nanonodule-decorated CNFs enhanced the quantity of the decorated WO3 on the CNFs surface. Hu and colleagues showed the electrospun preparation of mesoporous MnO2 and Mn3O4NPs-decorated CNFs, and observed that the hybrid CNFs have a diameter of 200-300 nm with high surface area. In another case, Xia and co-workers showed the overall synthesis of ultrafine transition metal oxide (Zn, Mn, and Co) NPs-embedded porous CNFs via a facile electrospinning strategy, following through the oxidisation method. As shown in Figure 4, there are plenty interconnected pores distributed within theZnO/CNFs, MnO/CNFs, and CoO/CNFs, and therefore the Zn, Mn, and Co elements are homogeneously distributed within the porous CNFs, respectively.

2.3.1 Zinc Oxide

Features a wide band gap of 3.37 eV that makes it a good nanomaterial in comparsion to others. ZnO may be a semiconducting material that exhibits high surface area to volume ratio, high biocompatibility, extremely stability, biomimetic, less toxicity and have a decent electron transferring feature. ZnO nanoparticles (NPs) are good supply for immobilization of proteins because of high



isoelectric point.Various biosensors with ZnO are for biomolecules like cholesterol, reported glucose, urea, cortisol, H2O2, glutamate and so on totally different shapes of ZnO were exploited to fabricate biosensors. For example, spherical and flower shaped ZnO NPs were immobilized on Au electrode to fabricate 2 cholesterolbiosensors (Umar et al., 2009a; Umar et al., 2009b). A layer of nafion was additionally applied to flower shaped ZnO. The sensorwith nation offered additional sensitivity and reproducibility, low km value, lower detection limit and fast response that were because of the network shaped by nafion which prevents protein leaking. Totally different structure of ZnO like ZnO nanorod and nanocom were also executed to construct glucose biosensor by modifying conductor (Wei et al., 2006; Wang et al., 2006). GOx was immobilized to each ZnO structure and nafion was used to cowl nanocombs. Presence of increase the enzyme-electrode nation will interaction and therefore increased performance of biosensor. 2 label free, low price immunosensors for detection of cortisol were made by using one dimensional ZnO nanorods (ZnO-NRs) and two dimensional ZnO nanoflakes (ZnO-NFs) (Vabbina et al., 2015). Both the nanostructures givemassive surface area, stability, biocompatibility and enhances the sensing ability of sensor. Theimmunosensors show selective electrochemical cortisol detection At 1 pM 100 times higher than enzyme connected immunosorbent assay (ELISA). 7.74 μ A/M and 11.86 μ A/M values were the sensitivity measured for ZnO-NFs and ZnO-NRs respectively. Kumar et al. (2015) showed а chemical biosensor for hydrazine that is ultrahighly sensitive and relies on ZnO nanocones. The sensor offered а high sensitive of $50 \times 10^4 \mu A/\mu M/cm^2$ and low detection limit of 0.01 μM.

ZnO usually shaped nanocomposite with polymers, AuNPs, graphene MWCNTs and so on and these hybrids are documented for immobilization of enzymes that helps in direct electron transfet. Between the conductor and active site which ends in superior properties for amperometric sensing. An amperometric glucose biosensor with great selectivity, stability, reliableness and repeatability was made-up by the use of the ZnO and MWCNTs on glassy carbon electrode (Palanisamy et al., 2012). The sensor exhibited a linear vary of 0.2-27.2 mm, 20 mM as limit of detection and a sensitivity of 4.18 mA/ mM. A ZnO/graphene and S6 aptamers primarily based biosensor for detection of SK-BR-3 breast cancer cells was made on ITO electrode (Liu et al., 2014). A low detection limit of 58 cells mL^{-1} and a

dynamic linear range of 10^2 - 10^6 cells mL⁻¹ was observed. Lately, extremely sensitive uric acid biosensor was developed with the use of ZnO nanosheets grown on electrode with high sensitivity, stability, duplicability and selectivity (Ahmad et al., 2015). ZnO nanosheets give additional surface to interact and lead to higher electron transfer between active sites of electrode and enzyme. Extent of communication between enzyme and electrode plays a key role in defining the level of sensitivity, i.e., more the interaction, higher the electron transfer and thus increased sensitivity.

III. APPLICATION OF THE NANOMATERIALS IN SENSORS

3.1 Gas sensors

Li and coworkers made CNFs of graphitic nanorolls employing а straightforward electrospinning-assisted solid-phase graphitization method, graphitic CNFs exhibit sensitivity to H2, CO, CH4, and ethanol gases at room temperature, and therefore the detection limit for CO gas is as low as 50 ppm. Zhang et al. reported a H2S detector using ZnO-CNFs composites, the asprepared H2S sensor showed a linear response within the limits of 50-102 ppm of H2S. Claramunt et al. deposited metal alloy NPs-decorated CNFs on Kapton for checking the level of NH3. The results show that the sensitivity of Au and Pd NPsdecorated CNFs to NH3 may be improved by dominant proportion of Au and Pd. Moreover, the latency of the detector is up to five minutes at 110-120 °C. However, when put next with the spectroscopical sensor like mid-infrared sensor and quartz-enhanced photoacoustic sensor, that have the benefits of fast detection at room temperature with no reagent, the operation temperature of Au, and Pd NPs-decorated CNFs is a lot higher. So as to scale down the detection temperature, Lee et al. changed WO3 nanonodules onto the CNFs, and the ready WO3 nanomodule-decorated CNFs not only provides a better sensing surface area, but also WO2+ on the surface of the fabric will mix with the O22 of NO2, realizing the detection of NO2 gas at room temperature, and the detection limit for NO2 reach 1 ppm.

3.2 Sensors for small molecules

CNFs-based nanomaterials being used to detect gas molecules and strain sensing, can also be used to detect micro molecules. Huang et al. loaded palladium NPs on CNFs to organize a Pd/CNFs modified carbon paste conductor for the detection of dopamine (DA), uric acid (UA), and ascorbic acid (AA). Once being modified with Pd NPs-



(Pd/CNFs). loaded **CNFs** the oxidation overpotentials of DA, UA, and AA were considerably reduced compared to the bare carbon paste electrode. The detection limits of Pd/CNFs changed carbon paste conductors for DA, UA and AA were 0.2 μ M, 0.7 μ M, and 15 μ M, respectively, and therefore the the} linear range was 0.5-160 µM (DA), 2-200 mm (UA), and 0.05-4 mm (AA). Liu et al. rumored another Pd NPsloaded CNFs modified carbon paste electrode for oxalic acid detection, had the detection limit of the sensor for oxalic acid as low as 0.2 mM, and shows a linear range from 0.2 to 45 nM. Liu et al. also made a Ni/CNFs composite electrode by electrospinning for glucose detection. The Ni/CNFs hybrid shows higher sensitivity towards glucose due to the electrocatalytic activity of the Ni NPs and therefore the stability of the carbon conductor. In the absence of chloride poisoning, the detection limit of the Ni/CNFs composite electrode for glucose is 1 μ M, with a linear range of 2 μ M–2.5 mm (R = 0.9997). Li and coworker successfully created а magnetic composite through polymerization of dopamine, laccase, and Ni NPs loaded CNFs. The as-prepared magnetic composite exhibited high selectivity towards catechol, and showed a linear range from 1 to 9100 μ M, with a detection limit of 0.69 µM for catechol in water samples. Lee et al. created a ZnO/CNFs composite for detecting DMMP, and ZnO NPs on CNFs enhanced the precise surface area of the detector and its affinity for DMMP. The detection limit of ZnO/CNFs composite for DMMP is 0.1 ppb, with a linear limits of 0.1–1000 ppb.

3.3 Biosensors

Ways to regulate and monitor health issues have attracted intensive attention, and even a lot of economic resources are designated to new technologies researches and development. The quicker the changes on the organism operate are detected, the simpler their control can be. Furthermore, identification systems with low detection limit, which implies more sensitivity, will increase the chances of medical intervention on unwellness treatment more efficiently. During this way, devices for biomolecules detection and quantification are of nice importance to boost life quality.

CNT field-effect transistors (CNTFET) have been with success enforced in dna sensors. Tran et al. rumored detection of influenza a virulent disease employing a CNTFET-based polymer sensor, achieving a limit of detection of 1 pM in a linear range from 1 pM to 10 nM. Dudina et al. reported the development of biosensors through an array of 9216 CNTFETs, detecting resistance within the 50 k Ω to 1 G Ω range, achieving a noise performance of 2.14 pARMS at a 1 kHz bandwidth and 0.84 pARMS at a 1 MHz bandwidth.

IV. LIMITATIONS

Carbon nanostructures have some limitations in their application due structural characteristics and properties. like functionalization. defects, particles and aggregation. All of these limitations are regarding sensor system responses and might decrease the standard of the device. This subject can present some limitations for the carbon nanostructures mentioned previously in this paper. CNT based sensor application is restricted due some of its structure characteristics. CNTs can not be applied directly in medical specialty applications due their metallic characteristic, being insoluble in several solvents. This characteristic can be a problem looking on the kind of device for biological concerns, that is being developed. Also, CNT sizes obtained from the synthesis sometimes aren't homogeneous, which may lead one to get results that are not consistent. CNT are often used as CNT-based biosensors operative with a field-effect transistor configuration (CNT-bioFET) to find biomolecules. Although this sort of system presents superior performance, the presence of background noise of electrostatic nature can phase out the information being measured. reliable As recognized by Camilli and Passacantando, the noise has identical scale range of the signal, and its origin is associated to substrate interactions and surface adsorbates. Another concern is that the presence of in CNT structure and even the defects functionalization methodology used for CNT application. They will influence CNT Young's modulus, which may yield many issues for sensor devices based on CNT specially those ones accustomed measure stress and strain properties.

V. CONCLUSION

Carbon nanostructure-based sensors are systems with excellent properties, giving us many prospects of application.Although carbon nanotubes have high reactivity just like the alternative allotropes, their mechanical properties conjointly imply within the development of sensors for physical parameter detection, like stress and strain. Their high surface-to-volume rate also permits the development of gas sensors nearly as good as those made using graphene structure. The devices reported throughout this review represent the utmost development found these days for carbon nanostructure-based sensors, presenting the



most analysis areas being explored and therefore the potentialities for the future. Several remained challenges regard on the carbon-based manufacture to terminate contaminants also as to improve the functionalization process. In addition to this, new synthesis methods for CNFs could be developed, also the biological modification of CNFs for subsequent biomedical applications including biosensors, anti-bacterial materials, bone tissue engineering, and others could be further explored. Improving these aspects can increase the sensor selectivity and manage defect densification, which is strictly regarding physical and chemical properties that has got to be overcome.

Conflict of interest

The authors declare no conflicts of interest.

REFERENCES

- [1]. Ng, S.M.; Koneswaran, M.; Narayanaswamy, R. A review on fluorescent inorganic nanoparticlesfor opticalsensing applications. RSC Adv. 2016, 6, 21624– 21661.
- [2]. Namdari, P.; Daraee, H.; Eatemadi, A. Recent advances in silicon nanowire biosensors: Synthesis methods, properties, and applications. Nanoscale Res. Lett. 2016, 11, 406.
- [3]. Wang, L.; Wu, A.G.; Wei, G. Graphenebased aptasensors: From molecule-interface interactions to sensordesign and biomedical diagnostics. Analyst 2018, 143, 1526–1543.
- [4]. Wang, L.; Zhang, Y.J.; Wu, A.G.; Wei, G. Designed graphene-peptide nanocomposites for biosensorapplications: A review. Anal. Chim. Acta 2017, 985, 24–40.
- [5]. Xia, Y.; Li, R.; Chen, R.S.; Wang, J.; Xiang, L. 3D architectured graphene/metal oxide hybrids for gas sensors: A review. Sensors 2018, 18, 1456.
- [6]. Hahm, J.I. Fundamental properties of onedimensional zinc oxide nanomaterials and implementations in various detection modes of enhanced biosensing. Annu. Rev. Phys. Chem. 2016, 67, 691–717.
- [7]. Zhao, Q.X.; Zhao, M.M.; Qiu, J.Q.; Lai, W.Y.; Pang, H.; Huang, W. One dimensional silver-based nanomaterials:Preparations and electrochemical applications. Small 2017, 13, 1701091.
- [8]. Li, W.; Zhang, F.; Dou, Y.Q.; Wu, Z.X.; Liu, H.J.; Qian, X.F.; Gu, D.; Xia, Y.Y.; Tu, B.; Zhao, D.Y. Aself-template strategy for the synthesis of mesoporous carbon

nanofibers as advanced supercapacitorelectrodes. Adv. Energy Mater. 2011, 1, 382–386.

- [9]. Barsan, M.M.; Ghica, M.E.; Brett, C.M.A. Electrochemical sensors and biosensors based on redoxpolymer/carbon nanotube modified electrodes: A review. Anal. Chim. Acta 2015, 881, 1–23.
- [10]. Meyyappan, M. Carbon nanotube-based chemical sensors. Small 2016, 12, 2118– 2129.
- [11]. Chen, L.F.; Lu, Y.; Yu, L.; Lou, X.W. Designed formation of hollow particle-based nitrogen-doped carbonnanofibers for highperformance supercapacitors. Energy Environ. Sci. 2017, 10, 1777–1783.
- [12]. Ning, P.G.; Duan, X.C.; Ju, X.K.; Lin, X.P.; Tong, X.B.; Pan, X.; Wang, T.H.; Li, Q.H. Facile synthesis of carbon nanofibers/MnO2 nanosheets as high-performance electrodes for asymmetric supercapacitors.Electrochim. Acta 2016, 210, 754–761.
- [13]. Teradal, N.L.; Jelinek, R. Carbon nanomaterials in biological studies and biomedicine. Adv. Healthc. Mater.2017, 6, 1700574.
- [14]. Abd El-Aziz, A.M.; El Backly, R.M.; Taha, N.A.; El-Maghraby, A.; Kandil, S.H. Preparation and characterizationof carbon nanofibrous/hydroxyapatite sheets for bone tissue engineering. Mater.Sci. Eng. C 2017, 76,1188–1195.
- [15]. Li, Y.; Zhang, M.F.; Zhang, X.P.; Xie, G.C.; Su, Z.Q.; Wei, G. Nanoporous carbon nanofibers decorated withplatinum nanoparticles for non-enzymatic electrochemical sensing of H2O2. Nanomaterials 2015, 5, 1891–1905.
- [16]. Magana, J.R.; Kolen'ko, Y.V.; Deepak, F.L.; Solans, C.; Shrestha, R.G.; Hill, J.P.; Ariga, K.; Shrestha, L.K.;Rodriguez-Abreu, C. From chromonic self-assembly to hollow carbon nanofibers: Efficient materials in supercapacitor and vapor-sensing applications. ACS Appl. Mater. Interfaces 2016, 8, 31231–31238.
- [17]. Chen, L.F.; Feng, Y.; Liang, H.W.; Wu, Z.Y.; Yu, S.H. Macroscopic-scale threedimensional carbon nanofiberarchitectures for electrochemical energy storage devices. Adv. Energy Mater. 2017, 7, 1700826.
- [18]. Shen, Y.; Li, L.; Xiao, K.J.; Xi, J.Y. Constructing three-dimensional hierarchical architectures by integratingcarbon nanofibers into graphite felts for water



purification. ACS Sustain. Chem. Eng. 2016, 4, 2351–2358.

- [19]. Zhang, L.F.; Aboagye, A.; Kelkar, A.; Lai, C.L.; Fong, H. A review: Carbon nanofibers from electrospunpolyacrylonitrile and their applications. J. Mater. Sci. 2014, 49, 463– 480.
- [20]. Feng, L.C.; Xie, N.; Zhong, J. Carbon nanofibers and their composites: A review of synthesizing, properties and applications. Materials 2014, 7, 3919–3945.
- [21]. Yue, Y.; Hu, G.Z.; Zheng, M.B.; Guo, Y.; Cao, J.M.; Shao, S.J. A mesoporous carbon nanofiber-modifiedpyrolytic graphite electrode used for the simultaneous determination of dopamine, uric acid, and ascorbicacid. Carbon 2012, 50, 107–114.
- [22]. Koehne, J.E.; Marsh, M.; Boakye, A.; Douglas, B.; Kim, I.Y.; Chang, S.Y.; Jang, D.P.; Bennet, K.E.; Kimble, C.;Andrews, R.; et al. Carbon nanofiber electrode array for electrochemical detection of dopamine using fastscan cyclic voltammetry. Analyst 2011, 136, 1802–1805.
- [23]. Rand, E.; Periyakaruppan, A.; Tanaka, Z.; Zhang, D.A.; Marsh, M.P.; Andrews, R.J.; Lee, K.H.; Chen, B.;Meyyappan, M.; Koehne, J.E. A carbon nanofiber based biosensor for simultaneous detection ofdopamineand serotonin in the presence of ascorbic acid. Biosens. Bioelectron. 2013, 42, 434–438.
- [24]. Periyakaruppan, A.; Gandhiraman, R.P.; Meyyappan, M.; Koehne, J.E. Label-free detection of cardiactroponin-I using carbon nanofiber based nanoelectrode arrays. Anal. Chem. 2013, 85, 3858–3863.
- [25]. Lee, J.S.; Kwon, O.S.; Park, S.J.; Park, E.Y.; You, S.A.; Yoon, H.; Jang, J. Fabrication of ultrafine metal-oxide-decorated carbon nanofibers for dmmp sensor application. ACS Nano 2011, 5, 7992–8001.
- [26]. Lee, J.S.; Kwon, O.S.; Shin, D.H.; Jang, J. Wo3 nanonodule-decorated hybrid carbon nanofibers for NO2 gas sensor application. J. Mater. Chem. A 2013, 1, 9099–9106.
- [27]. Hu, A.; Curran, C.; Tran, C.; Kapllani, A.; Kalra, V. Fabrication of transition metal oxide-carbon nanofibers with novel hierarchical architectures. J. Nanosci. Nanotechnol. 2014, 14, 5501–5507.
- [28]. Xia, G.L.; Zhang, L.J.; Fang, F.; Sun, D.L.; Guo, Z.P.; Liu, H.K.; Yu, X.B. General synthesis of transition metaloxide ultrafine nanoparticles embedded in hierarchically porous carbon nanofibers as advanced

electrodesfor lithium storage. Adv. Funct. Mater. 2016, 26, 6188–6196.

- [29]. Huang, Y.P.; Miao, Y.E.; Ji, S.S.; Tjiu, W.W.; Liu, T.X. Electrospun carbon nanofibers decorated with Ag-Ptbimetallic nanoparticles for selective detection of dopamine. ACS Appl. Mater. Interfaces 2014, 6, 12449–12456
- [30]. Huang, J.S.; Liu, Y.; You, T.Y. Carbon nanofiber based electrochemical biosensors: A review. Anal. Methods 2010, 2, 202–211.
- [31]. Li, W.; Zhang, L.S.; Wang, Q.; Yu, Y.; Chen, Z.; Cao, C.Y.; Song, W.G. Low-cost synthesis of graphitic carbonnanofibers as excellent room temperature sensors for explosive gases. J. Mater. Chem. 2012, 22, 15342–15347.
- [32]. Yang, C.; Denno, M.E.; Pyakurel, P.; Venton, B.J. Recent trends in carbon nanomaterial-based electrochemical sensors for biomolecules: A review. Anal. Chim. Acta 2015, 887, 17–37.
- [33]. Zhang, J.T.; Zhu, Z.J.; Chen, C.M.; Chen, Z.; Cai, M.Q.; Qu, B.H.; Wang, T.H.; Zhang, M. ZnO-carbonnanofibers for stable, high response, and selective H2S sensors. Nanotechnology 2018, 29, 275501.
- [34]. Petruci, J.F.D.; Fortes, P.R.; Kokoric, V.; Wilk, A.; Raimundo, I.M.; Cardoso, A.A.; Mizaikoff, B. Real-timemonitoring of ozone in air using substrate-integrated hollow waveguide mid-infrared sensors. Sci. Rep.2013, 3, 3174.
- [35]. Cho, E.; Perebikovsky, A.; Benice, O.; Holmberg, S.; Madou, M.; Ghazinejad, M. Rapid iodine sensing on mechanically treated carbon nanofibers. Sensors 2018, 18, 1486.
- [36]. Adabi, M.; Saber, R.; Faridi-Majidi, R.; Faridbod, F. Performance of electrodes synthesized with polyacrylonitrile-based carbon nanofibers for application in electrochemical sensors and biosensors. Mater. Sci. Eng. C 2015, 48, 673–678.
- [37]. Kosterev, A.; Wysocki, G.; Bakhirkin, Y.; So, S.; Lewicki, R.; Fraser, M.; Tittel, F.; Curl, R.F. Application of quantum cascade lasers to trace gas analysis. Appl. Phys. B 2008, 90, 165–176.
- [38]. Viciani, S.; de Cumis, M.S.; Borri, S.; Patimisco, P.; Sampaolo, A.; Scamarcio, G.; De Natale, P.; D'Amato, F.;Spagnolo, V. A quartz-enhanced photoacoustic sensor for H2S trace-gas detection at 2.6 μm. Appl. Phys. B 2015, 119, 21–27.



- [39]. Klocke, J.L.; Mangold, M.; Allmendinger, P.; Hugi, A.; Geiser, M.; Jouy, P.; Faist, J.; Kottke, T. Single-shot sub-microsecond mid-infrared spectroscopy on protein reactions with quantum cascade laser frequency combs. Anal. Chem. 2018, 90, 10494–10500.
- [40]. Brandstetter, M.; Lendl, B. Tunable midinfrared lasers in physical chemosensors towards the detection of physiologically relevant parameters in biofluids. Sens. Actuators B Chem. 2012, 170, 189–195.
- [41]. Galao, O.; Baeza, F.J.; Zornoza, E.; Garces, P. Strain and damage sensing properties on multifunctional cement composites with CNF admixture. Cem. Concr. Compos. 2014, 46, 90–98.
- [42]. Wu, Z.Y.; Li, C.; Liang, H.W.; Chen, J.F.; Yu, S.H. Ultralight, flexible, and fireresistant carbon nanofiber aerogels from bacterial cellulose. Angew. Chem. Int. Ed. 2013, 52, 2925–2929
- [43]. Eissa, S.; Alshehri, N.; Abduljabbar, M.; Rahman, A.M.A.; Dasouki, M.; Nizami, I.Y.; AlMuhaizea, M.A.; Zourob, M. Carbon nanofiber-based multiplexed immunosensor for the detection of survival motor neuron 1, cystic fibrosis transmembrane conductance regulator and duchenne muscular dystrophy proteins. Biosens. Bioelectron. 2018, 117, 84–90.
- [44]. Rizwan, M.; Koh, D.; Booth, M.A.; Ahmed, M.U. Combining a gold nanoparticlepolyethylene glycolglycol nanocomposite and carbon nanofiber electrodes to develop a highly sensitive salivary secretory immunoglobulin a immunosensor. Sens. Actuators B Chem. 2018, 255, 557–563.
- [45]. Arumugam, P.U.; Chen, H.; Siddiqui, S.; Weinrich, J.A.P.; Jejelowo, A.; Li, J.; Meyyappan, M. Wafer-scale fabrication of patterned carbon nanofiber nanoelectrode arrays: A route for development of multiplexed, ultrasensitive disposable biosensors. Biosens. Bioelectron. 2009, 24, 2818–2824.
- [46]. Gupta, R.K.; Periyakaruppan, A.; Meyyappan, M.; Koehne, J.E. Label-free detection of Creactive protein using a carbon nanofiber based biosensor. Biosens. Bioelectron. 2014, 59, 112–119.
- [47]. Palanisamy, S., Cheemalapati, S., Chen, S.-M., 2012. Int. J. Electrochem. Sci. 7, 8394– 8407
- [48]. W. Wei, J. Nong, Y. Zhu et al., "Graphene/Au-enhanced plastic clad silica

fiber optic surface plasmon resonance sensor," Plasmonics, vol. 13, no. 2, pp. 483–491, 2018.

- [49]. H. Jeong, D. M. Nguyen, M. S. Lee, H. G. Kim, S. C. Ko, and L. K. Kwac, "N-doped graphenecarbon nanotube hybrid networks attaching with gold nanoparticles for glucose non-enzymatic sensor," Materials Science and Engineering: C, vol. 90, pp. 38–45, 2018
- [50]. Y. Liu, A. Kannegulla, B. Wu, and L.-J. Cheng, "Quantum dot fullerene-based molecular beacon nanosensors for rapid, highly sensitive nucleic acid detection," ACS Applied Materials & Interfaces, vol. 10, no. 22, pp. 18524–18531, 2018.
- [51]. M.-J. Li, Y.-N. Zheng, W.-B. Liang, R. Yuan, and Y.-Q. Chai, "Using p-type PbS quantum dots to quench photocurrent of fullerene-Au NP@MoS2 composite structure for ultrasensitive photoelectrochemical detection of ATP," ACS Applied Materials & Interfaces, vol. 9, no. 48, pp. 42111–42120, 2017.
- [52]. Y. Hao, P. Yan, X. Zhang et al., "Ultrasensitive amperometric determination of PSA based on a signal amplification strategy using nanoflowers composed of single-strand DNA modified fullerene and Methylene Blue, and an improved surfaceinitiated enzymatic polymerization," Microchimica Acta, vol. 184, no. 11, pp. 4341–4349, 2017.
- [53]. C. Zhang, J. He, Y. Zhang et al., "Cerium dioxide-doped carboxyl fullerene as novel nanoprobe and catalyst in electrochemical biosensor for amperometric detection of the CYP2C19*2 allele in human serum," Biosensors and Bioelectronics, vol. 102, pp. 94–100, 2018.
- [54]. P. Chen, T. Wang, X. Zheng, D. Tian, F. Xia, and C. Zhou, "An ultrasensitive electrochemical immunosensor based on C60-modified polyamidoamine dendrimers and Au NPs for CO-catalytic silver deposition," New Journal of Chemistry, vol. 42, no. 6, pp. 4653–4660, 2018.
- [55]. O. Uygun, Ç. Şahin, M. Yılmaz, Y. Akçay, A. Akdemir, and F. Sağın, "Fullerene-PAMAM(G5) composite modified impedimetric biosensor to detect Fetuin-A in real blood samples," Analytical Biochemistry, vol. 542, pp. 11–15, 2018.
- [56]. Nag, M. Castro, V. Choudhary, and J.-F. Feller, "Sulfonated poly(ether ether ketone) [SPEEK] nanocomposites based on hybrid nanocarbons for the detection and



discrimination of some lung cancer VOC biomarkers," Journal of Materials Chemistry B, vol. 5, no. 2, pp. 348–359, 2017.

- [57]. Ö. Alver, C. Parlak, P. Ramasami, and M. Şenyel, "Interaction between doped C60 fullerenes and piperazine-2,3,5,6-tetraone: DFT simulation," Main Group Metal Chemistry, vol. 41, no. 3-4, pp. 63–66, 2018.
- [58]. Ö. Alver, C. Parlak, P. Ramasami, and M. Şenyel, "Density functional theory study on the adsorption of valproic acid to doped fullerenes," Main Group Metal Chemistry, vol. 41, no. 3-4, pp. 67–71, 2018.
- [59]. Ma, C. Yang, S. Zhu, J. Song, and Y. Fu, "A new nanomatrix based on functionalized fullerene and porous bimetallic nanoparticles for electrochemical chiral sensing," New Journal of Chemistry, vol. 42, no. 12, pp. 9801–9807, 2018.